

## Vitamin B<sub>12</sub> and BF<sub>3</sub>-etherate as catalysts in synthesis of some C<sub>4</sub>-C<sub>12</sub>-alkyl β-D-xylopyranosides

Zorica D Petrović\*, Dejan Andjelković & Aleksandra Spasojević

Department of Chemistry, Faculty of Science, University of Kragujevac, Radoje Domanovic 12, P. O. Box 60, 34000 Kragujevac, Serbia and Montenegro  
E-mail: zorica@kg.ac.yu

Received 26 October 2004; accepted (revised) 21 March 2005

Two methods are presented for synthesis of some C<sub>4</sub>-C<sub>12</sub>-alkyl β-D-xylopyranosides. The first method is the vitamin B<sub>12</sub>-catalyzed reaction of glycosylation of acetobromoxyllose with alkanols (ROH) (C<sub>4</sub>-C<sub>12</sub>). The reaction is carried out with 2 mol% of vitamin B<sub>12</sub>, with respect to xylosyl bromide, under argon at room temperature. Under these conditions peracetylated C<sub>4</sub>-C<sub>12</sub>-alkyl β-D-xylopyranosides are obtained. Following chromatographic purification these products are de-esterified with a mixture of methanol-triethylamine-water (2:1:1) to give corresponding alkyl β-D-xylopyranosides in 50-60% yield. In all cases 3,4-di-O-acetyl-D-xylal is obtained, as the product of reductive elimination of peracetylated xylosyl bromide (15-25%). The second method is synthesis of C<sub>4</sub>-C<sub>12</sub>-alkyl β-D-xylopyranosides performed by glycosylation of corresponding alkanols with tetra-O-acetyl β-D-xylopyranose in the presence of BF<sub>3</sub>-etherate, as a Lewis acid catalyst. This glycosylation proceeds in only moderate yield (45-55%), but simplicity of this method and avoidance of expensive heavy metal catalysts make such procedure attractive.

**Keywords:** Xylopyranosides, vitamin B<sub>12</sub>, glycosylation, cyanocobalamin

**IPC: Int.Cl. 7 C 07 C**

Cyanocob(III)alamin (vitamin B<sub>12</sub>) is a complex compound active in human beings which is named "pigment of life". Intracellular vitamin B<sub>12</sub> can be converted to the two coenzyme forms namely, methylcobalamin and 5'-deoxyadenosylcobalamin and promoted a series of biochemical transformations *in vivo*; these coenzymes are essential for cell growth and replication<sup>1,2</sup>. Cyanocob(III)alamin is also chiral and non-toxic reagent in organic chemistry which has been used as catalyst for various types of reactions<sup>3,4</sup>. The catalytically active species in these reactions is cob(I)alamin obtained by electrochemical or chemical reduction (with activated Zn dust or with NaBH<sub>4</sub>) of vitamin B<sub>12</sub> [Co(III) to Co(I)]. The cob(I)alamin, obtained in this way, is a very powerful nucleophile<sup>5</sup> and reacts rapidly with alkyl or alkenyl halides to form organocob(III)alamins (from kinetic studies Schrauzer and Deutch<sup>6</sup> concluded that the rate law corresponds to an S<sub>N</sub>2 type substitution at carbon). Finally, cleavage of the Co-C bond of this intermediate affords products of the following reactions: reductive C-C bond formation, reductive elimination, hydrogenation, oxidation and

substitution<sup>7,8</sup> independent of the structure of the substrates and reaction conditions (**Figure 1**).

### Results and Discussion

In recent years we have studied the reactions of organic halides catalyzed by organocob(III)alamin (vitamin B<sub>12</sub>)<sup>10-12</sup> and reactions like glycosylation of xylose peracetate<sup>13</sup>. In continuation of this investigation the behaviour of brominated β-D-xylose peracetate in the B<sub>12</sub>-catalyzed reaction of glycosylation has been examined. Reactions of glycosylation of mono- or disaccharides are particularly interesting in synthesis of biodegradable surfactants, detergents and emulsifiers<sup>9</sup>.

The reaction was started with 2 mol% of vitamin B<sub>12</sub>, with respect to xylosyl bromide **1**, and with NaBH<sub>4</sub>, as a reducing agent. Then xylosyl bromide, dissolved in the corresponding alkanol (ROH) **3**, was added to the reaction mixture under argon atmosphere at room temperature. The mechanism of this reaction<sup>14,15</sup> proceeds *via* a xylosecob(III)alamin intermediate **2** which with alkanols (ROH) **3a-h** (C<sub>4</sub>-C<sub>12</sub>) gave peracetylated alkyl β-D-xylopyrano-

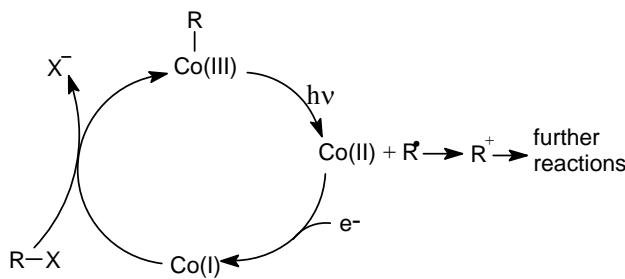


Figure 1

sides **4a-h**. Products **4a-h** were isolated by flash chromatography (yield 55-70%) and characterized by spectroscopic methods. Their <sup>1</sup>H and <sup>13</sup>C NMR spectral data were in agreement with structures assigned in previous work<sup>13</sup>. After de-esterification of **4a-h**, alkyl  $\beta$ -D-xylopyranosides **6a-h** were obtained in 50-60% yield (**Table I**). In all the cases 3,4-di-O-acetyl-D-xylal **5** was obtained (15-25%) as the product of reductive elimination of peracetylated xylosyl bromide (**Scheme I**). <sup>1</sup>H NMR characteristics of **5** were identical with those of a commercial sample.

Vitamin B<sub>12</sub>-catalyzed conversion of xylosyl bromide to alkyl  $\beta$ -D-xylopyranosides, described herein, illustrates that cob(I)alamin is a very strong nucleophile and cyanocob(III)alamin is powerful, natural and non-toxic catalyst which is now applied in complex carbohydrate chemistry.

For activation with a Lewis acid, exclusively the  $\beta$ -D-xylose tetraacetate **7** is required. The reaction with alkanols, ROH (C<sub>4</sub>-C<sub>12</sub>) **3a-h** and BF<sub>3</sub>-etherate, as a Lewis acid, gives corresponding *trans*-linked alkyl xylosides **4B**. Namely, under these conditions the presence of BF<sub>3</sub>-etherate leads to the formation of 1,2-acetoxonium intermediate<sup>13</sup> **8**. Addition of alcohol (ROH) at room temperature under anhydrous conditions gives *trans*-linked xylosides **4B**, as in previous reaction with SnCl<sub>4</sub> (ref. 13). The formation of the  $\beta$ -products **4** is kinetically controlled. Extension of reaction period leads to increasing proportions of the  $\alpha$ -xylosides **4a** due to anomerization. The acetylated alcohols (ROAc) are formed as side products in this reaction.

Following chromatographic purification the peracetylated products **4B** are de-esterified with sodium methoxide in anhydrous methanol (40 mL) under reflux and subsequently neutralized with ion-exchange resin (Amberlite IR 120 H<sup>+</sup>) to give alkyl  $\beta$ -D-xylosides **6a-h** in moderate yield (45-55%) (**Scheme II**).

**Table I** — Vitamin B<sub>12</sub> and BF<sub>3</sub>-etherate as catalysts in synthesis of some C<sub>4</sub>-C<sub>12</sub>-alkyl  $\beta$ -D-xylopyranosides **6a-h**

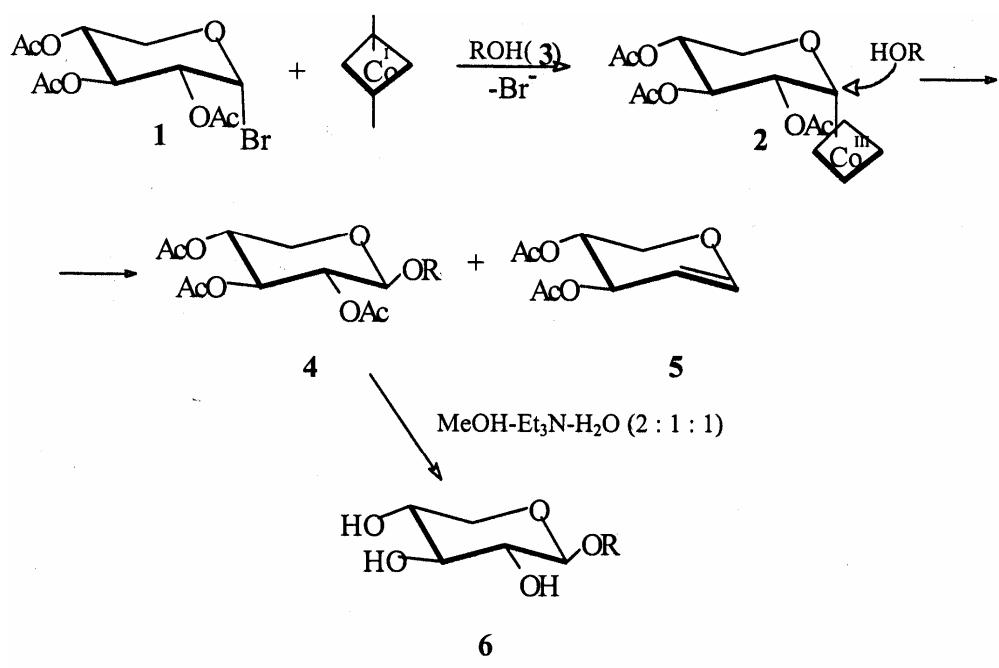
ROH (3)	R	Products <sup>a</sup>
<b>3a</b>	CH <sub>3</sub> (CH <sub>2</sub> ) <sub>3</sub>	<b>4a</b> <b>6a</b>
<b>3b</b>	CH <sub>3</sub> (CH <sub>2</sub> ) <sub>6</sub>	<b>4b</b> <b>6b</b>
<b>3c</b>	CH <sub>3</sub> (CH <sub>2</sub> ) <sub>7</sub>	<b>4c</b> <b>6c</b>
<b>3d</b>	CH <sub>3</sub> (CH <sub>2</sub> ) <sub>3</sub> CH(C <sub>2</sub> H <sub>5</sub> )CH <sub>3</sub>	<b>4d</b> <b>6d</b>
<b>3e</b>	CH <sub>3</sub> (CH <sub>2</sub> ) <sub>8</sub>	<b>4e</b> <b>6e</b>
<b>3f</b>	CH <sub>3</sub> (CH <sub>2</sub> ) <sub>9</sub>	<b>4f</b> <b>6f</b>
<b>3g</b>	CH <sub>3</sub> (CH <sub>2</sub> ) <sub>10</sub>	<b>4g</b> <b>6g</b>
<b>3h</b>	CH <sub>3</sub> (CH <sub>2</sub> ) <sub>11</sub>	<b>4h</b> <b>6h</b>

<sup>a</sup>Structures were determined on the basis of <sup>1</sup>H NMR and <sup>13</sup>C NMR spectral data<sup>13</sup>

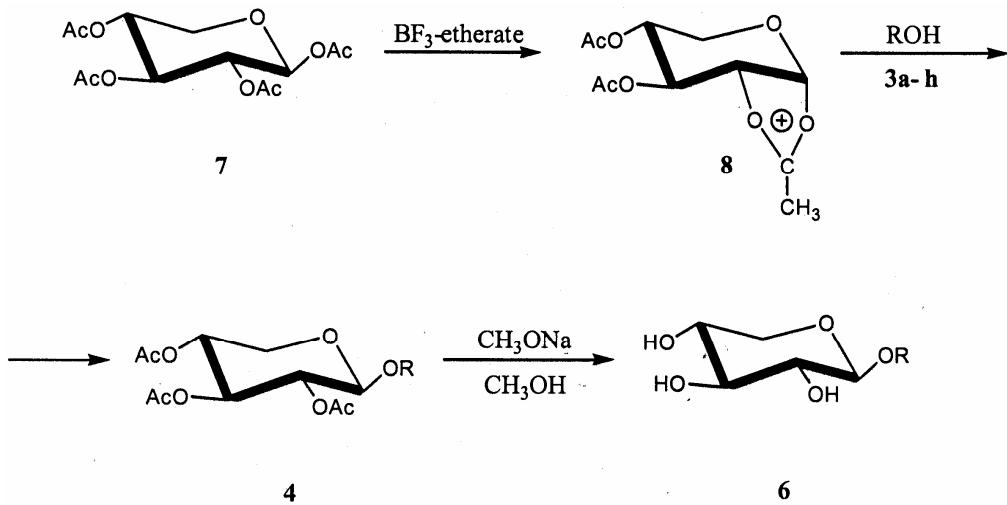
## Experimental Section

Flash column chromatography was carried out on Merc silica gel (particle size 0.04-0.06 mm) using dichloromethane and ethyl acetate as eluents. TLC was performed on Merck TLC aluminium sheets of silica gel 60 F<sub>254</sub>. Spots were visualized by spraying with 10% sulfuric acid in ethanol (carbohydrates) and with 1% anisaldehyde and 2% sulfuric acid in glacial acetic acid (non-carbohydrate compounds), respectively, and subsequent heating. <sup>1</sup>H and <sup>13</sup>C NMR spectra were run on a Varian Gemini 200 (200 MHz) or Bruker AC 250 (250 MHz) spectrometer in CDCl<sub>3</sub> with TMS as internal standard (chemical shifts in  $\delta$ , mol ppm). All solvents were purified by distillation.

**General procedure for preparation of alkyl xylosides catalyzed by vitamin B<sub>12</sub>.** A 100 mL flask, containing 20 mL of alkanol (ROH) **3** (C<sub>4</sub>-C<sub>12</sub>; **3a-h**), equipped with a magnetic stirrer bar under Ar, was charged with 0.16 g (0.12 mmole) of vitamin B<sub>12</sub> and 0.09 g (2.4 mmole) of NaBH<sub>4</sub>. After stirring for 30 min, the colour changed from red to dark green which indicates that Co(III) from vitamin B<sub>12</sub> is reduced to Co(I). Then tri-O-acetyl-D-xylopyranosyl bromide **1** (2.58 g, 8 mmole) (freshly prepared from peracetylated xylose **7** with hydrobromic acid in glacial acetic acid<sup>16,17</sup>) dissolved in 5 mL of alkanol (ROH), was added to this mixture. The colour immediately changed to red [xylose-Co(III)]. After stirring for 20 hr at room temperature, the dark red mixture was washed with cold saturated solution of NaCl and extracted with dichloromethane (3×25 mL). The combined organic phases were washed twice with water (2×25 mL), filtered over celite and evaporated



Scheme I



Scheme II

*in vacuo*. Most of the unreacted alkanol was removed by repeated evaporation of the syrup at 70°C on an oil pump in vacuum. The resulting syrup was purified by dry column chromatography (silica gel 60 (Merck) and dichloromethane-ethyl acetate (4:1) to give the products **4a-h** in the yield ranging from 55-70% (based on the peracetylated  $\beta$ -D-xylose) which were characterized by spectroscopic methods. In all the cases 3,4-di-O-acetyl-D-xylal **5** was obtained as the minor product (15-25%) as a result of reductive elimination of peracetylated xylosyl bromide **1**. Following chromatographic purification the peracetylated products **4a-h** were de-esterified with a

mixture of methanol-triethylamine-water (2:1:1) to give alkyl  $\beta$ -D-xylopyranosides **6a-h** in 50-60% yield.

**General procedure for preparation of alkyl xylosides in the presence of  $BF_3$ -etherate.** A solution of  $\beta$ -D-xylose tetraacetate (0.64 g, 2.01 mmoles) in anhydrous dichloromethane (20 mL) was stirred for 1-2 hr with molecular sieves (0.4 nm, 1-2 g) at room temperature and under argon atmosphere. The solution was treated with boron trifluoride etherate (4-20 mmoles) and then with the alcohol **3** (4-10 mmoles) dissolved in anhydrous dichloromethane (20 mL); the preparation of  $\beta$ -xylosides **4** required 2-3 days. Then the mixture was poured into saturated

NaHCO<sub>3</sub> solution (50 mL), the organic layer was separated and aqueous phase was extracted with dichloromethane (3×40 mL). The combined organic phases were washed twice with water (2×40 mL), filtered over Celite and evaporated *in vacuo*. Most of the unreacted alkanol was removed by repeated evaporation of the syrup at 70°C on an oil pump vacuum. Flash chromatography (toluene-ethyl acetate) of the resulting syrups afforded **4β** (yield 50-65%), which were characterized by spectroscopic methods. The acetylated alcohol was also isolated from the reaction mixture.

The resulting materials **4β** were deacetylated by treatment with sodium methoxide in anhydrous methanol (40 mL) under reflux and subsequently neutralized with ion-exchange resin (Amberlite IR 120 H<sup>+</sup>) to give alkyl  $\beta$ -xylosides **6a-h** in 45-55% yield.

## References

- 1 Retey J & Robinson J A, *Stereospecificity in Organic Chemistry and Enzymology*, (Veinheim), **1982**.
- 2 Branchaud B P & Friestad G K, *Vitamin B<sub>12</sub>*, edited by L Paquette (Wiley, New York), **1995**, 5511.
- 3 Scheffold R, Rytz G & Valder L, *Modern Synthetic Methods*, edited by R Scheffold (Wiley, New York), **1983**.
- 4 Pattenden G, *Chem Soc Rev*, **17**, **1988**, 361.
- 5 Das P K, O Hill H A, Pratt J M & Williams R J, *Biochim Biophys Acta*, **161**, **1967**, 646.
- 6 Schrauzer G N & Deutch E, *J Am Chem Soc*, **91**, **1969**, 3341.
- 7 Scheffold R, Abrecht S, Orlinski R, Ruf H R, Stamouli P, Tinembart O, Walder L & Weymuth C, *Pure Appl Chem*, **59**, **1987**, 363.
- 8 Shey J, McCauley K M, Dearth A, Young B & Van der Donk W A, *J Org Chem*, **67**, **2002**, 837.
- 9 Koeltzow D E & Urfer A D, *J Am Oil Chem Soc*, **61**, **1984**, 1651.
- 10 Petrovic Z D, Konstantinovic S, Scheffold R & Milosavljevic S, *Indian J Chem*, **36B**, **1999**, 765.
- 11 Petrovic Z, Bugarcic Z, Marjanovic Lj & Konstantinovic S, *J Mol Cat A: Chem*, **142**, **1999**, 393.
- 12 Petrovic Z, Mojsilovic B & Bugarcic Z, *J Mol Cat A: Chem*, **170**, **2001**, 267.
- 13 Konstantinovic S, Petrovic Z, Spasojevic A & Mojsilovic B, *Indian J Chem*, **40B**, **2001**, 614.
- 14 Forbes C L & Franck R W, *J Org Chem*, **64**, **1999**, 1424.
- 15 Mattheus de Roode B, Zuilhof H, Franssen M C R, Van der Padt A & Groot A, *J Chem Soc, Perkin Trans 2*, **2000**, 2217.
- 16 Rosevear P, Van Aken T, Baxter J & Ferguson-Miller S, *Biochemistry*, **19**, **1980**, 4108.
- 17 Kennedy J F & White C A, *Bioactive Carbohydrates in Chemistry, Biochemistry and Biology*, (Ellis Horwood Limited, Chichester), **1983**.